

larger decrease in the rate of hydride ion abstraction. Thus, there is a decrease in rate with increasing pH. Around neutral pH, the reaction proceeds mainly through path C in Scheme III. For the reasons stated earlier, there occurs a change in rate-limiting steps from the breakdown of the bromo ester (i.e., the k_2' step) to the disappearance of the imine (i.e., the k_3' and k_3'' steps). In the case of alanine and valine, the imine intermediate is converted into two sets of products by reacting with either water or by interacting with the oxidizing species. Of these, the reaction with water (i.e., the k_3' step) is an acid-catalyzed reaction and the other (i.e., the k_3'' step) is a base-catalyzed reaction. Hence, an increase in pH above the pK_a of the imine inhibits the reaction with water but enhances the interaction with the oxidizing species and causes a net increase in rate.

Conclusion

The mechanisms of oxidative decarboxylation of amino acids promoted by NBS have been shown to be significantly influenced by the presence of alkyl groups at the α -carbon. It will be interesting to see if this effect extends to other amino acids and to oxidizing agents other than NBS. It has been shown that the proton inventory technique can provide considerable information about the mechanisms of such oxidative decarboxylation reactions and its use in such studies should prove worthwhile.

Experimental Section

Materials. Glycine, glycine- d_5 , alanine, DL-valine, citric acid, acetic acid, acetic acid- d (98 atom % D), glycine ethyl ester hydrochloride, and betaine hydrochloride (99%) were all commercial samples and were used as obtained. *N*-Bromosuccinimide (Aldrich) was recrystallized from water. Deuterium oxide (99.75 atom % D; Bio-Rad) was used as obtained. Water was twice glass distilled before use.

Citric acid-disodium hydrogen phosphate buffer solutions of desired pH were prepared by mixing the required volumes of stock solutions of 0.1 M citric acid and 0.2 M disodium hydrogen phosphate.

Acetic acid-sodium acetate buffer solutions of the desired pH(D) were prepared by mixing the required volumes of stock solutions of acetic acid (or acetic acid- d) and sodium acetate in protium oxide or deuterium oxide. The ionic strength of the stock solutions was maintained with sodium perchlorate at 0.5 M. Solutions of lower buffer concentration were prepared from the above buffer solution by dilution with 0.25 M sodium perchlorate solution. The pH(D) of the solutions was measured by using a Corning pH meter Model 130 equipped with a combination electrode.

Kinetics. The oxidative decarboxylations of glycine, alanine, and valine were monitored by following the decrease in absorbance at 240 nm on a Cary 118C UV-vis spectrophotometer equipped with a constant-temperature cell compartment and cell holder to maintain a constant temperature.

Reactions were initiated by injecting 25 μ L of a stock solution of NBS in acetonitrile into 3.00 mL of the appropriate buffer solution containing the substrate. The runs were generally initiated exactly 75 s after the injection of the substrate solution. Reactions were followed to greater than 80% completion. Absorbance values at 5- or 10-s intervals were collected by using a Micromation computer interfaced to the Cary 118C spectrophotometer. The data were then analyzed by using a nonlinear least-squares computer program. Plots of $\log(A_t - A_\infty)$ vs. time were used in a confirmatory fashion.

The reactions of alanine or valine with NBS (6×10^{-4} M) at pH 3.7 (acetic acid-sodium acetate buffer) at 25 °C exhibited no dependence on amino acid concentration over the range 0.6×10^{-2} to 18×10^{-2} M. The first-order rate constants were 5.54×10^{-2} s $^{-1}$ and 1.54×10^{-2} s $^{-1}$, respectively, within experimental error. The concentration of NBS at pH 3.7 (ionic strength was 0.5 M) at 25 °C was varied from 1×10^{-4} to 8.4×10^{-4} M (glycine concentration was 1.22×10^{-2} M) and was shown to have no effect on the first-order rate constant for the oxidative decarboxylation of glycine ($k = 4.36 \times 10^{-3}$ s $^{-1}$ within experimental error). Variation of the ionic strength from 0 to 0.28 M (NaClO $_4$) at pH 5 did not influence the rate constant ($k = 8.43 \times 10^{-3}$ s $^{-1}$) for the glycine decarboxylation.

Acknowledgment. Support of this research by the Robert A. Welch Foundation is gratefully acknowledged.

Registry No. Glycine, 56-40-6; DL-alanine, 302-72-7; DL-valine, 516-06-3; *N*-bromosuccinimide, 128-08-5.

Carbonyl Methylenation Using a Titanium-Aluminum (Tebbe) Complex¹

Stanley H. Pine,* Robert J. Pettit, Gregory D. Geib, Susana G. Cruz,^{2a} Claudio H. Gallego,^{2b} Tomas Tijerina,^{2c} and Randall D. Pine

Department of Chemistry and Biochemistry, California State University, Los Angeles, Los Angeles, California 90032

Received November 6, 1984

The titanium-aluminum (Tebbe) complex [Cp $_2$ TiCH $_2$ ClAl(CH $_3$) $_2$] is shown to be an effective methylenating agent for a variety of carbonyl groups. The reaction is unique in that the carbonyl groups of carboxylic acid derivatives are readily methylenated. Thus vinyl enol ethers are prepared from esters and enamines are formed from amides. The complex provides a method for methylenating hindered or base sensitive ketones that is advantageous to the Wittig reagent. Selective methylenation of dicarbonyl compounds is also accomplished.

Methylenation and general alkylidenation of ketones and aldehydes using the Wittig reaction is well established in the methodology of organic synthesis.³ There are, however

some limitations to the use of the technique. Wittig reactions are quite sensitive to the steric environment around the carbonyl group undergoing reaction.⁴ Further, the

(1) We gratefully acknowledge support from the following: the donors of the Petroleum Research Fund, administered by the American Chemical Society; the NIH Minority Biomedical Research Support (MBRS) program; and the NIH MARC-Honors Undergraduate Research Training Program, NIGMS.

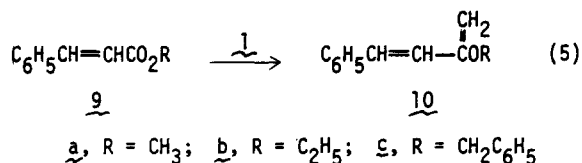
(2) (a) Participant in the NIH-MARC program. (b) Participant in the NIH-MBRS program. (c) Participant in the ACS-SEED program.

(3) (a) Maercker, A. *Org. React. (N.Y.)* 1965, 14, 270. (b) House, H. O. "Modern Synthetic Reactions"; Benjamin, Menlo Park, CA, 1972; pp 682-709. (c) Bestman, H. J.; Vostrowsky, O. *Top. Cur. Chem.* 1983, 109, 65.

(4) McMurry, J. E.; Choy, W. *Tetrahedron Lett.* 1980, 21, 2477. Boeckman, R. K., Jr.; Silver, S. M. *Ibid.* 1973, 3497. Sowerby, R. L.; Coates, R. M. *J. Am. Chem. Soc.* 1972, 94, 4758.

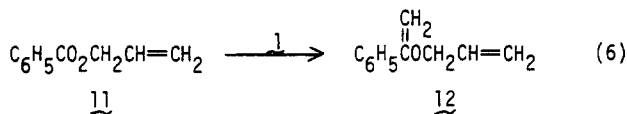
nium metallacycles from **1** plus alkene has been reported by Grubbs and co-workers.^{10b,17}

We were concerned that the reagent might interact competitively with the carbonyl and alkene double bonds. That has proven not to be a problem for substituted alkenes as demonstrated by the methylenations of unsaturated esters **9a-c** (eq 5). Furthermore, the stereochemical



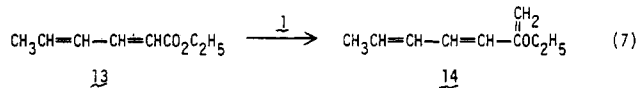
integrity of the carbon-carbon double bond is maintained during the reaction as demonstrated by using both the *Z* and *E* isomers of **9b**.⁸

Interestingly, methylenation of the allyl ester **11** leads to only a moderate yield of the enol ether **12** (eq 6) plus



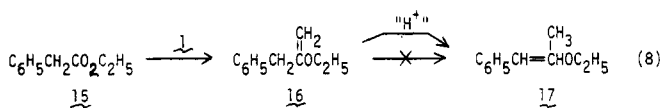
unreacted starting material. It appears that, in this case, **1** may interact with the terminal carbon-carbon double bond and is subsequently lost to the methylenation process.

The methylenation of ethyl sorbate (**13**) provides an example of the possible utility of the ester methylenation to produce a 2-substituted triene **14** (eq 7) or related

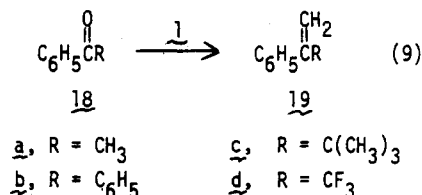


polyene, the potential precursor for an electrocyclic reaction. Stevenson and Bryson used such an approach to synthesize a 1,5-diene.¹⁸

The possibility was also investigated that the new carbon-carbon double bond might isomerize to a thermodynamically more stable position in the presence of the titanium and aluminum species from the complex. That was shown not to be a problem by using as examples the esters **7a** and **15**. In both cases, the more stable, conjugated product was not observed during the reaction. However, the conversion of **16** to **17** in the presence of a proton source did confirm the expected thermodynamic stabilities in that case (eq 8).

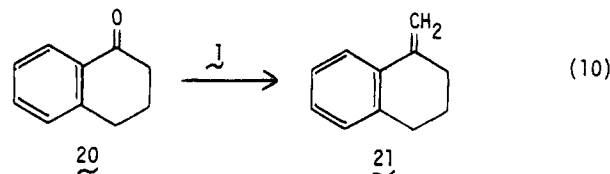


Ketones. Tebbe and co-workers reported the methylenation of cyclohexanone using **1**.⁷ Although the Wittig and a variety of other reactions are useful for ketone methylenation reactions,^{3,5d,19} the general importance of this conversion encouraged us to explore the possibility that complex **1** might complement those methods. A series of ketones (**18a-d**), dissolved in THF, toluene, or benzene was treated with **1** (eq 9). Methylenation was rapid in

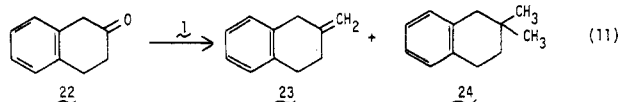


both the hydrocarbon and the Lewis base solvents, showing that ketone reactivity is greater than that of esters, a result similar to the well-established reactions of the carbonyl group with nucleophiles. Of interest is the observation that reactions proceed well with methyl **18a** as well as *tert*-butyl **18c** ketones.

Methylenation of 1-tetralone (**20**) (eq 10) proceeds with no problems from the peri interactions or enolization.

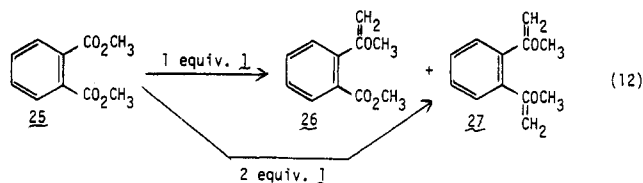


Methylenation of 2-tetralone (**22**) illustrates an interesting competitive reaction (eq 11). Reaction of equimolar



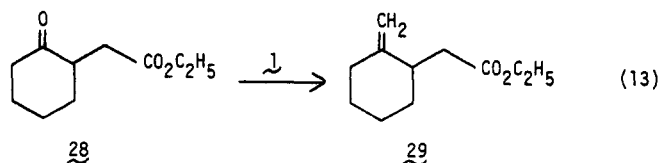
quantities of **22** and **1** leads to the expected methylenation product **23** along with 20–25% of **24**. The dimethylation product presumably results from the reaction of **23** with **1**. Grubbs and co-workers have observed related results in their work with titanium metallacycles.^{10b,20}

Dicarbonyl Compounds. The possibility that selective methylenation might occur using **1** was also explored. Thus, the diester, dimethyl phthalate (**25**), when treated with 1 equiv of **1** yields monomethylenation product **26** along with 20–30% of dimethylenation product **27** (eq 12).



The result reflects an approximate statistical methylenation of equivalent ester groups. When 2 equiv of complex are used, dimethylenation takes place to give only **27**.

The observation that ketone carbonyl groups react with **1** considerably faster than do ester carbonyl groups suggested that it might be possible to selectively methylenate only one carbonyl in a keto ester. Thus when **28** is treated with **1**, only **29**, the product of ketone methylenation, is observed (eq 13).



We have similarly found that a 1,2-keto ester (a pyruvate) can undergo selective mono- or dimethylenation using

(20) Grubbs, R. H., unpublished results.

(17) Howard, T. R.; Lee, J. B.; Grubbs, R. H. *J. Am. Chem. Soc.* **1980**, *102*, 6876.

(18) Stevenson, J. W. S.; Bryson, T. A. *Tetrahedron Lett.* **1982**, *23*, 3143.

(19) Eisch, J. J.; Piotrowski, A. *Tetrahedron Lett.* **1983**, *24*, 2043. Johnson, C. R.; Elliot, R. C.; *J. Am. Chem. Soc.* **1982**, *104*, 7041. Corey, E. J.; Kang, J. *Ibid.* **1982**, *104*, 4724. Fieser, M.; Fieser, L. F. "Reagents for Organic Synthesis"; Wiley: New York, 1975; Vol. 5, p 458, 527.

1-Phenyl-1-(3-propenoxy)ethene (12): method A, 50%; ¹H NMR (CCl₄) δ 4.15 (d, 1, *J* = 3 Hz), 4.4 (m, 2), 4.65 (d, 1, *J* = 3 Hz), 5.1-5.6 (m, 2), 5.8-6.3 (m, 1), 6.2-6.8 (m, 5). Anal. (C₁₁H₁₂O) C, H.

2-Ethoxy-1,3,5-heptatriene (14): method A, 54%; ¹H NMR (CCl₄) δ 1.35 (t, 3, *J* = 7 Hz), 1.75 (d, 3, *J* = 5 Hz), 3.75 (q, 2, *J* = 7 Hz), 3.95 (s, 2), 5.4-6.6 (m, 4). Anal. (C₉H₁₄O) C, H.

2-Ethoxy-3-phenylpropene (16):⁸ method A, 90%; ¹H NMR (CDCl₃) δ 1.2 (t, 3, *J* = 8 Hz), 3.3 (s, 2), 3.7 (q, 2, *J* = 8 Hz), 3.8 (d, 1, *J* = 2 Hz), 3.9 (d, 1, *J* = 2 Hz), 7.2 (s, 5).

2-Ethoxy-1-phenylpropene (17):²⁴ from isomerization of 16 in CDCl₃ to give 83% of a mixture of 70% *E* and 30% *Z* isomers; ¹H NMR (CDCl₃) δ 1.3 (t, 3, *J* = 7 Hz), 1.9 (s, 3), 3.75 (q, 2, *J* = 7 Hz), 5.2 (s, 0.3), 5.5 (s, 0.7), 6.8-7.3 (m, 5).

1-Methyl-1-phenylethene (19a):²⁵ method A, 88%; ¹H NMR (CCl₄) δ 2.05 (s, 3), 4.95 (br s, 1), 5.25 (br s, 1), 7.0-7.4 (m, 5).

1,1-Diphenylethene (19b):²⁶ method A, 97%; ¹H NMR (CCl₄) δ 5.3 (s, 2), 7.1 (s, 10).

3,3-Dimethyl-2-phenyl-1-butene (19c): method A, 96%; ¹H NMR (CCl₄) δ 1.1 (s, 9), 4.65 (d, 1, *J* = 2 Hz), 5.05 (d, 1, *J* = 2 Hz), 6.8-7.1 (m, 5). Anal. (C₁₂H₁₆) mass spectrum.

3,3,3-Trifluoro-2-phenylpropene (19d): method A, 50%; ¹H NMR (CCl₄) δ 5.6 (d, 1, *J* = 1 Hz), 5.8 (d, 1, *J* = 1 Hz), 7.2 (s, 5). Anal. (C₉H₇F₃) mass spectrum.

1-Methylene-1,2,3,4-tetrahydronaphthalene (21):²⁷ method A, 73%; ¹H NMR (CCl₄) δ 1.8 (m, 2), 2.5 (m, 2), 2.8 (t, 2, *J* = 6 Hz), 4.8 (d, 1, *J* = 1 Hz), 5.3 (d, 1, *J* = 1 Hz), 6.9 (m, 3), 7.5 (m, 1).

2-Methylene-1,2,3,4-tetrahydronaphthalene (23):²⁸ method A, 40%, separated from 24 via chromatography; ¹H NMR (CCl₄) δ 2.45 (t, 2, *J* = 7 Hz), 2.85 (t, 2, *J* = 7 Hz), 3.5 (s, 2), 4.75 (br s, 2), 6.9 (s, 4).

2,2-Dimethyl-1,2,3,4-tetrahydronaphthalene (24):²⁹ method A, 15%, separated from 23 via chromatography; ¹H NMR (CCl₄) δ 0.95 (s, 6), 1.5 (t, 2, *J* = 6 Hz), 2.45 (s, 2), 2.7 (t, 2, *J* = 6 Hz), 6.8 (br s, 4).

Methyl 2-(1-methoxyethenyl)benzoate (26): method A, 45%, separated from 27 via chromatography; ¹H NMR (CCl₄) δ 3.55 (s, 3), 3.7 (s, 3), 4.1 (d, 1, *J* = 2 Hz), 4.2 (d, 1, *J* = 2 Hz), 7.0-7.4 (m, 3), 7.4-7.6 (m, 1). Anal. (C₁₁H₁₂O₃) C, H.

1,2-Bis(1-methoxyethenyl)benzene (27): method A, 65%, using 2 equiv of 1; ¹H NMR (CCl₄) δ 3.5 (s, 6), 4.1 (d, 2, *J* = 2

Hz), 4.2 (d, 2, *J* = 2 Hz), 6.9-7.3 (m, 4). Anal. (C₁₂H₁₄O₂) C, H.

Ethyl 2-(2-methylenecyclohexyl)acetate (29): method A, 67%; ¹H NMR (CCl₄) δ 1.15 (t, 3, *J* = 7 Hz), 1.4-2.6 (m, 11), 4.0 (q, 2, *J* = 7 Hz), 4.4 (s, 1), 4.5 (s, 1). Anal. (C₁₁H₁₈O₂) mass spectrum.

Methyl 2-phenyl-2-propenoate (31): method A, 72%; ¹H NMR (CCl₄) δ 3.7 (s, 3), 5.7 (d, 1, *J* = 1 Hz), 6.2 (d, 1, *J* = 1 Hz) 7.1-7.4 (m, 5). Anal. (C₁₀H₁₀O₂) mass spectrum.

2-Methoxy-3-phenyl-1,3-butadiene (32): method A, 45%; ¹H NMR (CCl₄) δ 3.6 (s, 3), 4.0 (d, 1, *J* = 2 Hz), 4.1 (br s, 1), 5.05 (br s, 1), 5.55 (d, 1, *J* = 2 Hz), 7.1 (s, 5). Anal. (C₁₁H₁₂O) mass spectrum.

1-(1-Phenylethenyl)piperidine (33):³⁰ method B, D, 76% (by NMR); ¹H NMR (CCl₄) δ 1.6 (m, 6), 2.8 (m, 4), 4.0 (s, 1), 4.1 (s, 1), 7.1-7.4 (m, 5). Anal. (C₁₃H₁₇N) mass spectrum.

4-(1-Phenylethenyl)morpholine (34):³¹ method D, 67% (by NMR); ¹H NMR (C₆D₆) δ 2.8 (m, 4), 3.7 (m, 4), 4.3 (s, 1), 4.6 (s, 1), 7.0-7.8 (m, 5). Anal. (C₁₂H₁₅NO) mass spectrum.

1-(1-Phenylethenyl)pyrrolidine (35):³⁰ method D, 80% (by NMR); ¹H NMR (C₆D₆) δ 1.7 (m, 4), 3.1 (m, 4), 4.2 (s, 1), 4.35 (s, 1), 7.1-7.8 (m, 5).

***N*-Methyl-*N*-2-propenylbenzenamine (36):**³² method D, 97% (by NMR); ¹H NMR (C₆D₆) δ 1.9 (s, 3), 3.0, (s, 3), 4.3 (s, 1), 4.35 (s, 1), 6.9-7.4 (m, 5).

***N*-*N*-Dimethyl-2-propenamine (37):**³² method D; ¹H NMR (C₆D₆) δ 1.9 (s, 3), 2.6 (s, 6), 3.85 (s, 1), 3.95 (s, 1).

Registry No. 1, 67719-69-1; 2a, 93-58-3; 3b, 99-75-2; 3c, 121-98-2; 3d, 1126-46-1; 4a, 4747-13-1; 4b, 51440-57-4; 4c, 51440-56-3; 4d, 67471-39-0; 5a, 939-48-0; 5b, 120-51-4; 5c, 93-99-2; 5d, 774-65-2; 6a, 42237-98-9; 6b, 25109-98-2; 6c, 19928-57-5; 6d, 78386-38-6; 7a, 553-86-6; 7b, 119-84-6; 8a, 74104-12-4; 8b, 74104-13-5; 9a, 103-26-4; 9b, 103-36-6; 9c, 103-41-3; 10a, 95045-97-9; 10b, 77882-39-4; 10c, 95045-98-0; 11, 583-04-0; 12, 40815-73-4; 13, 2396-84-1; 14, 95045-99-1; 15, 101-97-3; 16, 74104-11-3; (*E*)-17, 71094-33-2; (*Z*)-17, 71094-47-8; 18a, 98-86-2; 18b, 119-61-9; 18c, 938-16-9; 18d, 434-45-7; 19a, 98-83-9; 19b, 530-48-3; 19c, 5676-29-9; 19d, 384-64-5; 20, 529-34-0; 21, 25108-63-8; 22, 530-93-8; 23, 66448-77-9; 24, 13556-55-3; 25, 131-11-3; 26, 51440-54-1; 27, 95046-00-7; 28, 24731-17-7; 29, 53544-45-9; 30, 15206-55-0; 31, 1865-29-8; 32, 95046-01-8; 33, 14990-66-0; 34, 7196-01-2; 35, 3433-56-5; 36, 21267-55-0; 37, 22499-75-8; *N*-methylacetanilide, 579-10-2; 4-benzoylmorpholine, 1468-28-6; 1-benzoylpyrrolidine, 3389-54-6; dimethylacetamide, 127-19-5; 1-benzoylpiperidine, 776-75-0.

(30) von Hirsch, H. *Chem. Ber.* 1967, 100, 1289.

(31) Noyori, R.; Yokoyama, K.; Hayakawa, Y. *Org. Synth.* 1978, 58, 56.

(32) Ahlbrecht, H.; Raab, W. *Synthesis* 1980, 320.

(24) Blondeau, D.; Sliwa, H. *J. Chem. Res., Miniprint* 1979, 117.

(25) Bhacca, N. S.; Johnson, L. F.; Shooley, J. N. "NMR Spectra Catalog"; Varian Associates, Palo Alto, 1962, No. 232.

(26) Allen, C. F. H.; Converse, S. "Organic Syntheses; Wiley, New York, 1932; Vol. I, p 226.

(27) Meyers, A. I.; Ford, M. E. *J. Org. Chem.* 1976, 41, 1735.

(28) Morrison, H.; Giachero, D. *J. Org. Chem.* 1982, 47, 1058.

(29) Khalaf, A. A.; Roberts, R. M. *J. Org. Chem.* 1972, 37, 4227.

Solvent Effects on Keto-Enol Equilibria: Tests of Quantitative Models

Sander G. Mills and Peter Beak*

Department of Chemistry, University of Illinois, Urbana, Illinois 61801

Received June 26, 1984

The effect of solvent on the equilibrium constants between selected keto-enol isomer pairs has been determined. The protomeric systems studied are 5,5-dimethyl-1,3-cyclohexanedione (1), 2-methyl-1,3-cyclohexanedione (3), 3-oxabicyclo[4.3.0]-2,9-dioxonane (5), 2,4-pentanedione (7), ethyl 3-oxobutanoate (9), and 9-anthracenone (11). A wide variety of theoretical and empirical solvation parameters have been tested in a multiparameter linear free energy format to model the changes in equilibria. The most successful correlations are obtained with the Kamlet-Taft polarity-polarizability and hydrogen-bonding terms, although the Swain *A* and *B* factors have advantages in some cases. In general, for the isomer pairs in which the enol cannot form an internal hydrogen bond (1-2, 3-4, and 11-12), the equilibria appear to be controlled almost completely by the hydrogen-bonding basicity of the solvent. For the isomer pairs 5-6, 7-8, and 9-10, in which intramolecular hydrogen bonding is possible, the polarity-polarizability effect dominates, although differential stabilization of the isomers by hydrogen bonding remains significant.

The effect of solvent on the relative energies of hydroxy-pyridine-pyridone and mercaptopyridine-thio-

pyridone isomers is quantitatively correlated by an approach that models the differential interactions of the